

Plant Genomic DNA Purification Kit

Cat # : PT-PGDNA Size : 50 Reactions

Components of the kit :

	50 reaction	notice
*Solution E1	19 ml	Precipitate can be easily dissolved by incubating at 50°C.
*Solution E2 powder	0.23 g x 2	Each tube add 1 ml sterile water to dissolve. Store the solution E2 at -20°C.
* RNase A	10 mg	Store at 4°C when received the kit. Add 200 ul sterile water before use.
Solution P1	7.5 ml	
* Binding Buffer	12.5 ml	Add 25 ml Ethanol before using
* Wash buffer	12.5 ml	Add 50 ml Ethanol before using
Elution Buffer	7.5 ml	Preheated the solution at 60-70°C
Filter column & collection tube	50 sets	With blue ring
Binding column & collection tube	50 sets	

Special Note:

Sample size and yield:

Up to 100 mg for wet leaf weight, 50 mg for fungi.
Typical yield: 1-20 ug

Materials to be supplied by the user:

For tissue grinding: mortar and pestle.
Liquid nitrogen
Ethanol

Before starting:

Thaw the solution E2 before grinding tissue.
Preheat water bath or heating block at 65°C.

Step	Procedures
Sample Preparation	<ol style="list-style-type: none"> 1. Add liquid nitrogen and plant tissue (<100 mg) in mortar and pestle, grind the sample into fine powder. Do not let sample to thaw. 2. Immediately transfer the powder to 1.5 ml tube, add 360 µl Solution E1 , 40µl Solution E2 and 4 µl RNaseA Solution. Vortex vigorously for 5-10 sec.
Cell Lysis and removing proteins, polysaccharides	<ol style="list-style-type: none"> 3. Incubating at 65°C in water-bath or heating block for 20 min. Mix by inverting the tube from time to time. 4. Add 130µl Solution P1, mix by inverting the tube, and incubate on ice for 5 min. 5. Centrifuge at top speed (12-14,000 rpm) for 5 min at RT. 6. Collect the supernatant into Filter Column (With blue ring) sitting in a clean 2 ml collection tube. Spin at top speed for 2 min. 7. Collect the flow-through buffer in the collection tube into a new 1.5 ml tube, avoid pipetting the pellets.
Binding DNA	<ol style="list-style-type: none"> 8. Add 700µl of Binding Buffer and mix thoroughly by pipetting. 9. Transfer 650 µl of the mixture including any precipitate into Binding Column and spin for 1 min at top speed and discard the flow-through. 10. Repeat step 9 with remaining mixture.
Wash DNA	<ol style="list-style-type: none"> 11. Discard the flow-through, wash by adding 600 µl of Wash Buffer to the column, then spin at top speed (12-14,000 rpm) for 1 min. 12. Discard the flow-through and repeat this wash step. 13. Discard the flow-through, then centrifuge for 5 min at top speed (12-14,000 rpm) to remove ethanol.
Eluting DNA	<ol style="list-style-type: none"> 14. Discard the collection tube and place the column into a new 1.5 ml centrifuge tube. 15. Add preheated 60-70°C of 50-100 µl Elution Buffer or H₂O (pH should be between 7.0-8.5) into centre of the column, wait for 2 min. 16. Eluting the DNA by centrifugation for 2 min, discard the column and store the DNA at -20°C. <i>* Repeat elution once by using the same eluate and the same column may have 10-15% more DNA yield.</i>